



Research Article

Oxidative stress and toxicological impacts of Monomehypo exposure on bone marrow and erythrocytes in male Japanese quail

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ABSTRACT

Pesticide use in agriculture, particularly in developing countries, continues to pose a significant threat to avian populations. To investigate the potential harm caused by monomehypo pesticide, an experiment was conducted focusing on its effects on bone marrow and erythrocyte morphology in male Japanese quail. Forty-eight sexually mature quail weighing 120g were selected and divided into four groups (A, B, C and D), each comprising 12 quail. After a two-week adaptation period in their natural environment, the trial commenced. Group A was assigned as the control group, whereas groups B, C, and D were subjected to monomehypo at doses of 120µg/kg BW, 160µg/kg BW, and 200µg/kg BW respectively. The results demonstrated a significant decrease in peroxidase (POD), superoxide (SOD), catalase (CAT) and reduced glutathione (GSH) levels with an increase in dosage. Conversely, oxidative stress markers such as reactive oxygen species (ROS) and thiobarbituric acid reactive substances (TBARS) showed a significant increase in all the exposed groups. This study also analyzed morphological changes in erythrocytes, revealing various abnormalities such as blebbed nuclei, micronucleated cells, notched nucleated cells, condensed nucleated cells, lobed nuclei cells, nucleus remnants, and cells lacking cytoplasm. These alterations were observed across all experimental groups and intensified with both higher doses and longer exposure. The results indicated that exposure to monomehypo showed a toxic effect and caused minor to major alterations in male Japanese quail. These results emphasize the urgent need to address the detrimental impacts of pesticides on avian populations, particularly in developing countries where birds are disproportionately affected.

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Introduction

Living organisms are commonly subjected to an extensive range of synthetic and naturally derived substances. The industrial revolution and the subsequent advancements in synthetic chemistry have led to the creation of numerous novel chemicals, including fertilizers and pesticides (Osman et al. 2023). Approximately two million tons of pesticides are manufactured and utilized

annually worldwide. The United States consumes 24% of this total, while European countries account for 45% of pesticide usage. The remaining 25% is distributed among other regions globally. The most commonly employed pesticides, namely Lindane, dichlorodiphenyltrichloroethane (DDT), and malathion, make up 70% of the overall pesticide use (Rajak et al. 2023). Toxic pesticides are increasingly targeting a wide range of organisms. Fruits and

vegetable crops alone consume 27% of the total pesticide consumption (Umapathi et al. 2022). Insecticides comprise 80% of pesticide use, herbicides make up 15%, plant fungal diseases account for 1.46%, and the remaining 3% consists of other forms of pesticides. However, on a global scale, herbicide consumption stands at 47.5%, insecticides at 29.5%, fungicides at 17.5%, and the remaining 5.5% represents other types of pesticides. India is responsible for 76% of the worldwide pesticide consumption, compared to the global usage rate of 44% (Cestonaro et al. 2022). The presence of pesticides in ecosystems has detrimental effects, which vary depending on the concentration, quantity, and duration of exposure to the contaminants (Voltz et al. 2022). Pesticides exhibit rapid lethality when orally exposed. Despite their widespread popularity and use, mixing, applying, and treating plants and animals with pesticides pose serious health risks (Rashid et al. 2022). Monomehyppo, a systemic insecticide with contact and stomach action, disrupts nerve transmission in insects and binds to receptors in the nervous system. It belongs to the Carbamate group, specifically the 3rd age group, and is available in G Formulation (Granules) with a concentration of 5%w/w. Monomehyppo is particularly effective against *Chilo suppressalis* walker and *Tryporyza incertulas* walker. Furthermore, dithiocarbamate pesticides and carbamate compounds are known to induce oxidative stress through the augmentation of ROS production (Suliman et al. 2020), leading to endoplasmic reticulum interference and mitochondrial dysfunction (Li et al. 2022). These chemicals also impair leukocyte development and interfere with the particular function of immune cells (Li et al. 2021). Experimental evidence has shown that carbamate pesticides increase the risk of liver, kidney, thyroid, bladder, uterus, bone, brain, and adrenal gland tumors in humans (Mdeni et al. 2022). Epidemiological studies have confirmed the damaging impacts of pesticide exposure on various organs, including the liver, brain, colon, and lungs. Previous investigations have also revealed the fatal consequences of insecticides, such as cancer, in different individuals (Sharma et al. 2020). All pesticides possess a certain level of toxicity. Even products with low toxicity can cause health problems if animals are exposed to sufficient quantities. The risk of health issues depends not only on the toxicity of the ingredients but also on the level of exposure to the product. Avian species are highly sensitive to industrial chemicals and pesticides, making them valuable model organisms for assessing chemical toxicity in ecotoxicology. Birds can be exposed to toxic substances directly through food or dermal contact during preening and grooming. Oral intake is considered the most significant route of exposure for avian species. Therefore, conducting oral

toxicity tests is crucial in determining the toxicological effects of any compound under investigation on avian species (Djekkoun et al. 2021).

Materials and Methods

Experimental Design

The experiment was conducted in the research laboratory of the Department of Zoology at Islamia University of Bahawalpur, Pakistan. All the birds were reared in wire cages for a duration of 45 days. In this present study, 48 sexually mature Japanese quail weighing 120g were utilized. The quail were divided into four groups, namely A, B, C, and D, with 12 quail in each group. Throughout the experimental duration, the birds were provided with a corn-soybean meal-based feed containing 22% protein and water twice a day. Monomehyppo was administered orally to groups B, C, and D at doses of 120 µg/kg BW, 160µg/kg BW, and 200µg/kg BW, respectively, for a period of 45 days. Group A served as the control group. The quails were monitored on a daily basis, and samples were collected on the day 15th, 30th, and 45th day. The birds were housed at room temperature with a humidity level of 60-65%. The quails were maintained under a 12-hrs light/dark cycle and provided continuous ad libitum access to food and water throughout the duration of the experiment.

Blood Sampling and Morphological Changes in Erythrocytes

Blood sample (5 ml) was collected from jugular vein of each quail at day 15th, 30th, and 45th of the trail. For morphological and nuclear changes in red blood cells of exposed quail, fine thin blood films were prepared from fresh blood of each quail without any anticoagulants. All the blood films were immediately dried, fixed with absolute alcohol and stained with Giemsa solution for 5 to 10 secs. A computer-assisted light microscope was used to observe and score nuclear changes and micronucleus frequencies in a total of 3000 erythrocytes from each quail. The observations were conducted under an oil immersion lens (100X) (Cimrin et al. 2023).

Oxidative and Antioxidant Study in Bone Marrow

On the 15th, 30th, and 45th day of the trial, bone marrow samples were collected from each quail in every group. The collected samples were preserved in serum cups for subsequent procedures. Various oxidative enzymes were determined according to previous protocols including ROS (Miladinovic et al. 2021), TBARS (Tarladgis et al. 1960), and GSH (Owens and Belcher 1965). The status of various antioxidant enzymes was measured according to previous protocols including POD (Owens and Belcher 1965), SOD (Al-Matubsi et al. 2011), and

CAT (Goth. 1991) in the bone marrow of exposed quail.

Statistical analysis

Obtained data were statistically subjected to one-way analysis of variance (ANOVA) using IBM, SPSS statistics version 20. Post hoc-Tukey’s test was employed to compare the means of the treatments at a significance level of $P < 0.05$.

Results

Morphological and Nuclear Alterations in Erythrocytes

The formation of micronuclei in erythrocytes was observed as the most prominent abnormality in the monomethylhypoxanthine-exposed quail in groups B to D (120-200 µg/kg BW) at experimental days 15, 30, and 45. There was a significant increase in erythrocytes with lobed nuclei, blebbed nuclei, and notched erythrocytes in group D compared to group C. Throughout the 45-day period, there was a significantly increased in all treatment groups (B, C, and D) compared to the control group (A). At various time points, there was a significant increase

in the occurrence of lobed nucleated cells in exposed groups B and D compared to group A and C. Exposed groups B and C exhibited a significant increase in nucleus ruminants in red blood cells compared to the control group A at various time intervals. The presence of blebbed nucleated cells significantly increased in treatment group B at all-time points compared to the control group A. Additionally, in comparison to the control group, exposed group D exhibited a significant increase in blebbed nucleated.

Furthermore, exposed groups B, C and D demonstrated a significantly elevation in the occurrence of condensed nucleated cells as compared to unexposed group A throughout the experimental period. The absence of cytoplasm in cells significantly increased in treatment group D compared to the control group A at different time intervals. Treatment groups B, C and D displayed a significant increase in notched nucleated cells compared to the control group A over the 45-day period. Fig. 1 represents the all morphological and nuclear changes in erythrocytes of exposed quail.

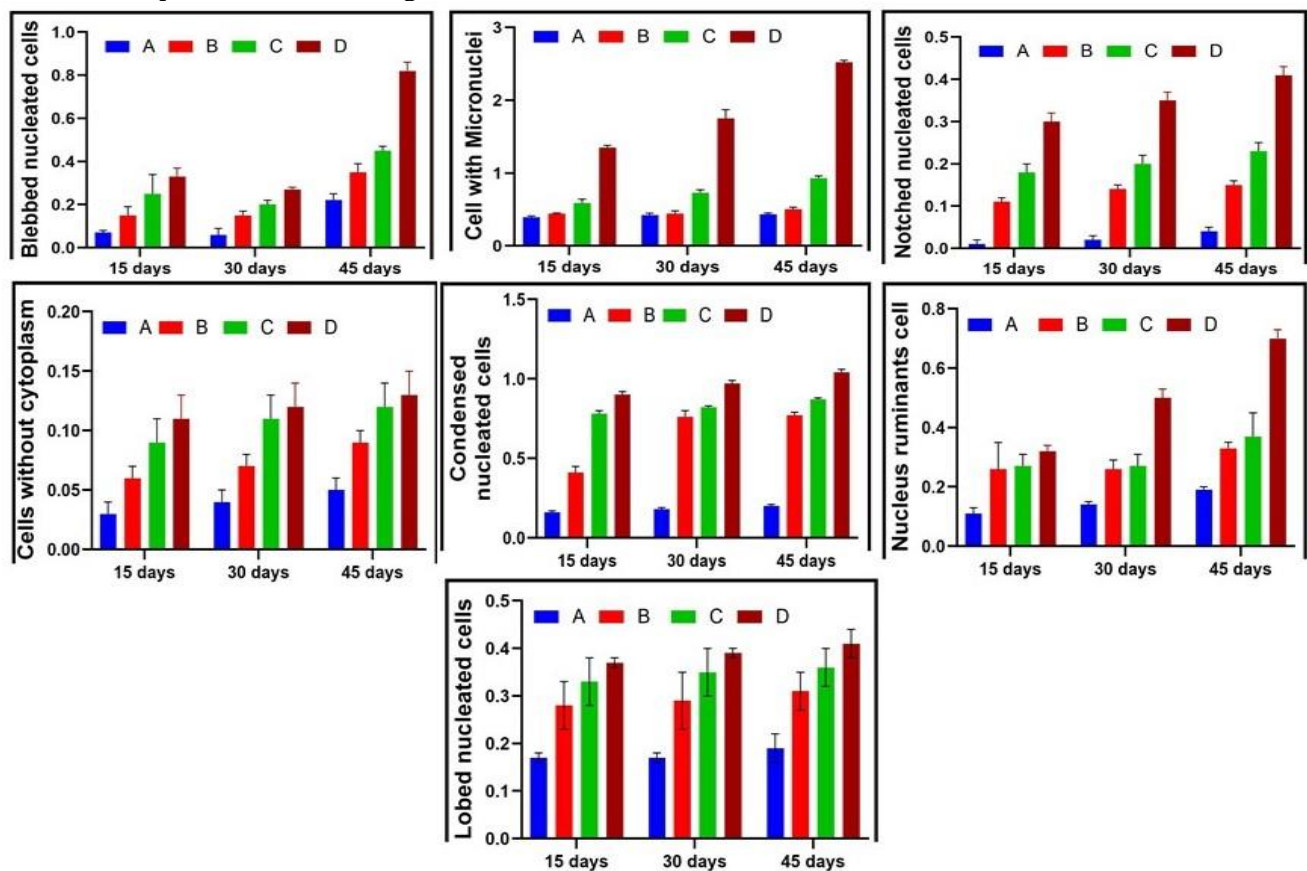


Fig. 1: Effects of Monomethylhypoxanthine exposure on erythrocyte (morphological and nuclear ailments) in male Japanese quail

Oxidative and Antioxidant Status in Bone Marrow

In terms of oxidative stress markers and antioxidant status, this study investigated the activity of ROS, TBARS, GSH, POD, SOD, and CAT (Fig. 2). The results revealed that monomethylhypochlorite exposure had a slightly significant effect on ROS activity after 30 days of treatment, with group D exhibiting a significant increase compared to the control group. This suggests that monomethylhypochlorite may contribute to the generation of ROS, leading to oxidative damage in the quail. Additionally, the activity of TBARS, an indicator of lipid peroxidation, showed a slightly significant effect after 30 days of monomethylhypochlorite exposure, with group D displaying a significant increase compared to the control group. This indicates that monomethylhypochlorite may induce lipid peroxidation and cause oxidative damage to cell membranes.

Furthermore, the activity of GSH, an important antioxidant defense molecule, demonstrated a

slightly significant effect after 45 days of monomethylhypochlorite exposure, with group D exhibiting a significant decrease compared to the control group. These findings suggest that monomethylhypochlorite might interfere with the antioxidant defense system, resulting in a reduction in GSH levels. In terms of the antioxidant enzymes evaluated (POD, SOD, and CAT), no significant changes in activity were observed after 15, 30, and 45 days of monomethylhypochlorite exposure in the control groups. However, a significant decrease in activity was observed in all the groups exposed to monomethylhypochlorite, indicating that monomethylhypochlorite may impact the activity of antioxidant enzymes in male Japanese quail. Moreover, the correlation analysis of all these biomarkers showed significant differences in all the groups when compared with the control group (Fig. 3).

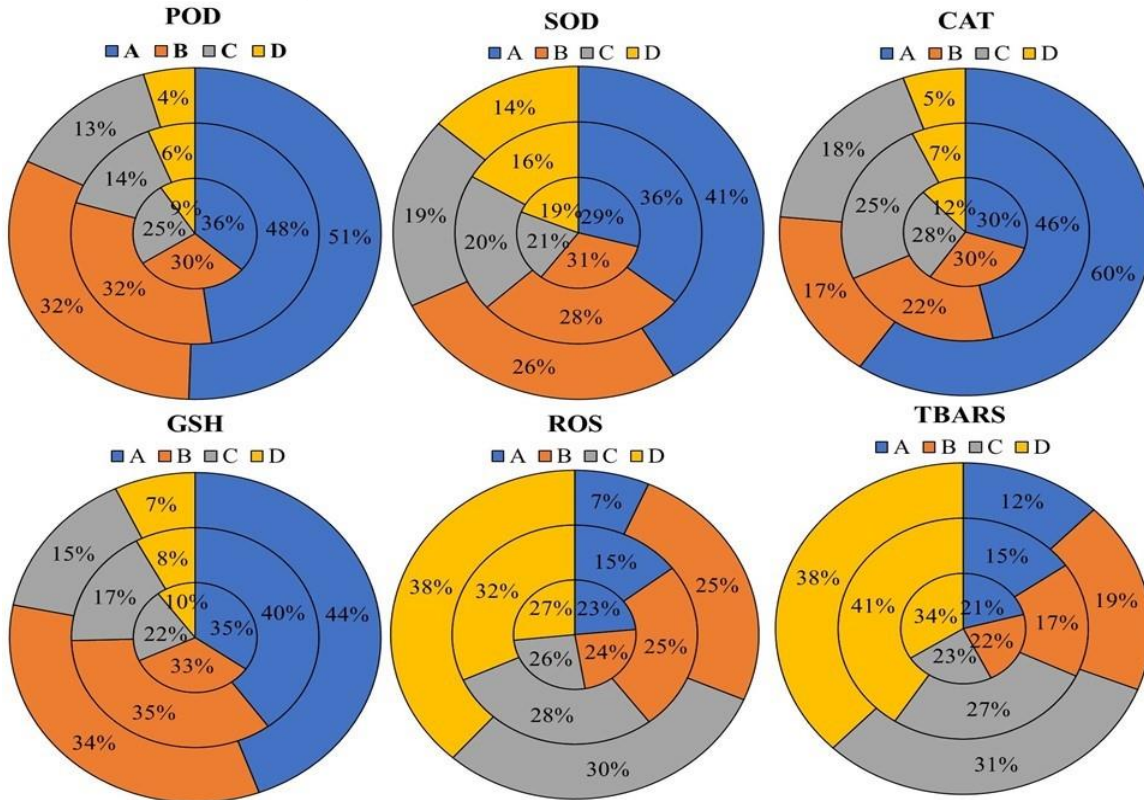


Fig. 2: Oxidative and antioxidant status in bone marrow of male Japanese quail exposed to various concentration on monomethylhypochlorite

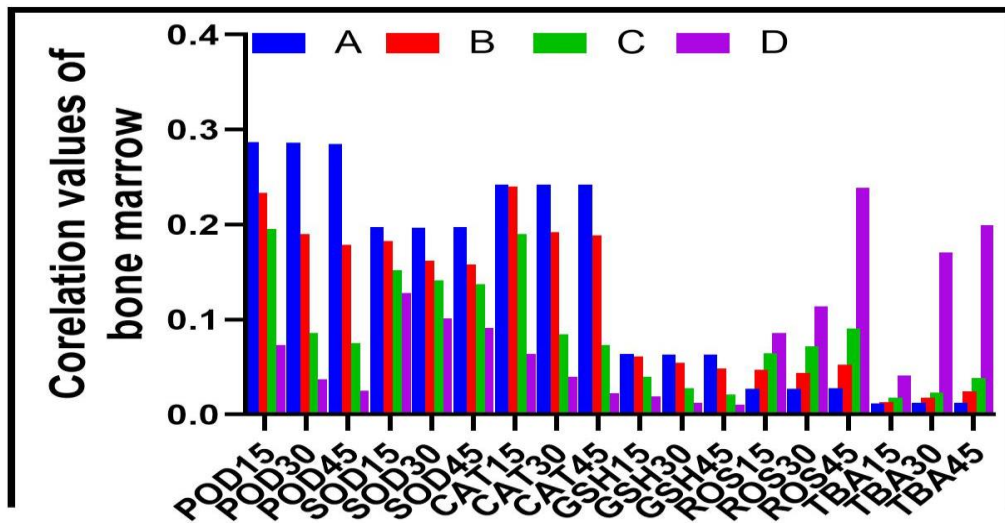


Fig. 3: Correlation analysis of biochemical analysis in male Japanese quail

Discussion

Monomethyl hypochlorite is a neurotoxic pesticide that causes toxicity in birds, mammals and aquatic species. According to reports, insecticides and pesticides can cause DNA damage that manifests as the production of micronuclei, chromosomal aberrations, and microtubule abnormalities (Kocaman and Topaktaş 2010; Hussain et al. 2015; Suljevic et al. 2019). The presence of micronuclei in the cytoplasm is thought to be a biomarker of DNA damage (Saleh and Sarhan 2007). Micronuclei are tiny, distinct nuclei that are 1/3rd the diameter and 1/10th in length of the main nucleus. They have the same colour, refraction, and texture as the nucleus. When the whole or chromosomal segments are not integrated in the primary nucleus following cell division, monomethyl hypochlorite treatment may lead to micronuclei (Hussain et al. 2012). Micronuclei in RBC arise from genetic damage, such as chromosomal damage, which leads to the lagging of fragments during anaphase or the failure of cytoplasmic chromatin-containing or acentric chromosomes to be properly incorporated into daughter nuclei (Ghaffar et al. 2015).

In our study, small satellite nuclei were observed that are independent from the cell's primary nucleus seen in the cytoplasm. The creation of cytoplasmic chromatin-containing entities that cannot be integrated into daughter nuclei (clastogenesis) due to genetic damage is frequently connected with micronuclei and following acentric chromosomes during anaphase (Khosrow et al. 2019). The micronuclei observed in our study exhibited consistent color and texture. Our findings are in aligned with (Quero et al. 2016), who also reported a positive correlation between the

frequencies of micronuclei and notched nuclei in parrots. These abnormalities were simultaneously observed affecting the same nucleus in three different cells.

In our present study, inconsistent but significant morphological changes were observed in the erythrocytes in a time and dose dependent manner. The changes in structure of RBC may be attributed to oxidative stress on the mitochondria, leading to the potential release of cytochrome c from the voltage dependent anion channels in the outer mitochondrial membrane. This, in turn, triggers apoptotic changes such as the cleavage of cytoskeletal proteins like gelsolin and fodrin, as well as increased production of caspase activated DNase in the nucleus of RBC, responsible for the deterioration and division of nuclear lamins proteins. Similar findings were reported by Hussain et al. (2012), who determined that exposure to ATZ (atrazine) lowered intracellular ATP level and mitochondrial membrane potential in a variety of cells, leading to both morphology and respiratory function. The frequency of formation of micronuclei increased in erythrocytes of male Japanese quail that were subjected to higher doses of monomethyl hypochlorite. As well as the micronuclei formation increased with the increase in exposure time. These findings align with previously published research. Our study also revealed decreased level of antioxidant parameters such as enzymes GSH, CAT, SOD, POD, and increased in oxidative stress, increased production of ROS, and TBARS, in the bone marrow of male Japanese quail. These findings are consistent with previous studies that have demonstrated the toxic effects of pesticides on quail (Arslan et al. 2022; Taha et al. 2013; Taha et

al. 2020). These results suggest that exposure to monomehypo can induce oxidative stress by altering the antioxidant enzyme status in the bone marrow and causing morphological changes in erythrocytes in male Japanese quail.

Conclusion

This current study demonstrates that monomehypo pesticide exposure has detrimental effects on male Japanese quail, particularly in terms of genotoxicity, cytotoxicity, and oxidative stress. The observed morphological alterations in erythrocytes, such as micronuclei formation and changes in nucleus shape, highlight the potential damage caused by monomehypo. The increased levels of ROS and lipid peroxidation, coupled with decreased antioxidant enzyme activity, further support the induction of oxidative stress by monomehypo. Overall, this study contributes to our understanding of the harmful effects of monomehypo pesticide on avian health and underscores the importance of implementing sustainable and eco-friendly agricultural practices to mitigate the potential risks to bird populations.

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Competing Interest:

The authors declare that they have no relevant financial or non-financial interests to disclose.

Author Contribution:

All authors contributed to the conception, design, and analysis of the study.

References

Al-Matubsi HY, Nasrat NA, Oriquat GA, Abu-Samak M, Al-Mzain KA and Salim M, 2011. The hypocholesterolemic and antioxidative effect of dietary diosgenin and chromium chloride supplementation on high-cholesterol fed Japanese quail. *Pakistan Journal of Biological Sciences* 14:425-432.

Arslan AS, Seven I, Mutlu SI, Arkali G, Birben N and Seven PT, 2022. Potential ameliorative effect of dietary quercetin against lead-induced oxidative stress, biochemical changes, and apoptosis in laying Japanese quail. *Ecotoxicology and Environmental Safety* 231:113200.

Akram R, Iqbal R, Hussain R, Jabeen F and Ali M, 2021. Evaluation of oxidative stress, antioxidant enzymes and genotoxic potential of bisphenol A in fresh water bighead carp (*Aristichthys nobilis*) fish at low concentrations. *Environmental Pollution* 268:115896.

Cestonaro LV, Macedo SMD, Piton YV, Garcia SC and Arbo MD, 2022. Toxic effects of pesticides on cellular and humoral immunity: an overview. *Immunopharmacology and Immunotoxicology* 44(6): 816-831.

Ghaffar A, Hussain R, Khan A, Abbas RZ, Aslam S, Mehreen M and Rani K, 2015. Hemato-biochemical and testicular changes induced by subchronic doses of triazophos in male Japanese quail. *Pakistan Journal of Agricultural Sciences* 52:801-807.

Goth L, 1991. A simple method for determination of serum catalase activity and revision of reference range. *Clinica Chimica Acta* 196:143-151.

Hussain R, Mahmood F and Khan A, 2015. Genotoxic and pathological effects of malathion in male Japanese quail (*Coturnix japonica*). *Pakistan Journal of Agricultural Sciences* 52:1149-1156.

Hussain R, Mahmood F, Khan A, Javed MT, Rehan S and Mehdi T, 2012. Cellular and biochemical effects induced by atrazine on blood of male Japanese quail (*Coturnix japonica*). *Pesticide Biochemistry and Physiology* 103:38-42.

Khosrow KY, Ali FE and Sadeq WS, 2019. Study of Genotoxic and Cytotoxic effects of Malathion in Japanese quail. *Tikrit Journal of Pure Science* 23:19-26.

Kocaman AY and Topaktas M, 2010. Genotoxic effects of a particular mixture of acetamiprid and α -cypermethrin on chromosome aberration, sister chromatid exchange, and micronucleus formation in human peripheral blood lymphocytes. *Environmental Toxicology: An International Journal* 25:157-168.

Mdeni NL, Adeniji AO, Okoh AI and Okoh OO, 2022. Analytical evaluation of carbamate and organophosphate pesticides in human and environmental matrices: a review. *Molecules* 27:618.

Miladinovic DC, Crnic AP, Pekovic S, Dacic S, Ivanovic S, Santibanez JF, Cupic V, Borozan N, Miljakovic EA and Borozan S, 2021. Recovery of brain cholinesterases and effect on parameters of oxidative stress and apoptosis in quail (*Coturnix japonica*) after chlorpyrifos and vitamin B1 administration. *Chemico-Biological Interactions* 333:109312.

Osman KA, Shaaban MM and Ahmed NS, 2023. Biomarkers of imidacloprid toxicity in Japanese quail, *Coturnix coturnix japonica*. *Environmental Science and Pollution Research* 30(3): 5662-5676.

Owens CWI and Belcher RV, 1965. A colorimetric micro-method for the determination of glutathione. *Biochemical Journal* 94:705.

Quero AA, Ferre DM, Zarco A, Cuervo PF and Gorla NB, 2016. Erythrocyte micronucleus cytome assay of 17 wild bird species from the central Monte desert, Argentina. *Environmental*

- Science and Pollution Research 23:25224-25231.
- Rajak P, Roy S, Ganguly A, Mandi M, Dutta A, Das K, Nanda S, Ghanty S and Biswas G, 2023. Agricultural pesticides–Friends or foes to biosphere? Journal of Hazardous Materials Advances 10:100264.
- Rashid S, Rashid W, Tulcan RXS and Huang H, 2022. Use, exposure, and environmental impacts of pesticides in Pakistan: a critical review. Environmental Science and Pollution Research 29:43675-43689.
- Saleh K and Sarhan MA, 2007. Clastogenic analysis of chicken farms using micronucleus test in peripheral blood. Journal of Applied Sciences Research 3:1646-1649.
- Sharma A, Shukla A, Attri K, Kumar M, Kumar P, Sutte A, Singh G, Barnwal RP and Singla N, 2020. Global trends in pesticides: A looming threat and viable alternatives. Ecotoxicology and Environmental Safety 201:110812.
- Suliman Khan A, Shah SSA, Gulfam N, Khisroon M and Zahoor M, 2020. Toxicity evaluation of pesticide chlorpyrifos in male Japanese quail (*Coturnix japonica*). Environmental Science and Pollution Research 27:25353-25362.
- Suljevic D, Corbic A, Islamagic E, Focak M, Filipic F and Alijagic A, 2019. Impairments of bone marrow hematopoietic cells followed by the sever erythrocyte damage and necrotic liver as the outcome of chronic in vivo exposure to cadmium: novel insights from quail. Environmental Toxicology and Pharmacology 72:103250.
- Taha AT and Douri AA, 2013. The effect of supplementing different levels of fenugreek seeds (*Trigonella foenum-graecum*) on physiological performance of Japanese quail (*Coturnix-cutornix japonica*) exposed to oxidative stress induced by hydrogen peroxide. Pharmacognosy Communications 3:26.
- Taha AT, Al-Jumaily TK and Al-Samrai MK, 2020. August. Effect of melatonin in adult quail males exposed to oxidative stress induced by H₂O₂. In IOP Conference Series: Earth and Environmental Science 553:012013.
- Tarladgis BG, Watts BM, Younathan MT and Dugan JrL, 1960. A distillation method for the quantitative determination of malonaldehyde in rancid foods. Journal of the American Oil Chemists' Society 37:44-48.
- Umapathi R, Ghoreishian SM, Sonwal S, Rani GM and Huh YS, 2022. Portable electrochemical sensing methodologies for on-site detection of pesticide residues in fruits and vegetables. Coordination Chemistry Reviews 453:214305.
- Voltz M, Guibaud G, Dages C, Douzals JP, Guibal R, Grimbuhler S, Grunberger O, Lissalde S, Mazella, N, Samouëlian A and Simon S, 2022. Pesticide and agro-ecological transition: assessing the environmental and human impacts of pesticides and limiting their use. Environmental Science and Pollution Research 291:1-5.